Use of cholesterol and soluble tumour markers CEA and syndecan-2 in pleural effusions in cases of inconclusive cytology

Miklos Gulyas, Janos Fillinger, Andras D Kaposi, Miklos Molnar

ABSTRACT

Aims In order to improve diagnostics in pleural effusions, additional value of effusion cholesterol, carcinoembryonic antigen (CEA) and syndecan-2 assays to cytology was studied.

Methods Biomarkers were measured in effusion supernatants from 247 patients, of whom 126 had malignant pleural involvement, and their additional diagnostic efficacy to cytology was assessed.

Results Syndecan-2 measurement, although gave detectable concentrations in all effusions with highest median value in mesotheliomas, was non-discriminative between different pathological conditions. CEA concentrations exceeding 5 ng/mL cut-off point indicated carcinomas, regardless of pleural involvement, which gave a sensitivity of 62% and specificity of 100% for carcinoma. Cholesterol concentration over 1.21 mmol/L cut-off value indicated neoplastic pleural involvement with 99% sensitivity and ‘merely’ 69% specificity, the latter mainly due to raised levels being associated also with benign inflammatory effusions. Combined CEA and cholesterol determinations increased the sensitivity for diagnosing carcinomatosis from 70% with cytology alone to 84% and established the correct diagnosis in 16 of 31 carcinomatosis cases with inconclusive cytology. Cholesterol measurement alone, with elevated level, in combination with absence of substantial number of inflammatory cells in effusion sediment proved to be a magnificent marker for neoplastic pleural involvement with 99% efficacy, and recognised all 36 such cases with inconclusive cytology.

Conclusions Simultaneous measurement of CEA and cholesterol concentrations in effusion, or at least cholesterol alone, in combination with non-inflammatory fluid cytology, provides additional specific information about neoplastic pleural involvement, and can therefore be used as an adjunct to cytology, above all, in inconclusive cases.

INTRODUCTION

Tumour diagnosis in pleural fluid is based primarily on effusion cytology. However, conventional cytological examination of pleural effusion many times gives inconclusive result: beside malignant and benign diagnoses, numerous cases are equivocal, ‘suspicious for malignancy’. In pleural irritations caused by inflammation, liver cirrhosis, irradiation and chemotherapy it is often difficult to discriminate shed reactive mesothelial cells from well differentiated cancer cells. In addition, cytology is frequently false negative, as result of low effusion volume with small yield and/or poor preservation of tumour cells. Consequently, a specific cytological diagnosis may have a sensitivity of only 43%–80%.

Samples which are inconclusive due to some cellular atypia can be investigated further by auxiliary morphological techniques, such as immunocytochemistry and DNA cytometry, although these methods are of little help when only a small number of tumour cells available conserved.

Biochemical indicators, such as concentrations of cholesterol and carcinoembryonic antigen (CEA) in the serous effusions, have been studied as markers of malignancy. Such tests can be carried out in adequately stored fluids after cytological examination quickly. In a previous study we investigated the contribution of ascitic fluid levels of cholesterol and CEA to cytology. In the thoracic cavity, compared with the peritoneal cavity, other cancers and benign conditions are prevalent, for example, lung cancers, mesothelioma, tuberculosis and purulent inflammations versus gastrointestinal tumours, liver cirrhosis and pancreatitis, which may affect cut-off values and diagnostic performance.

Syndecan-2 (fibroglycan) is a cell surface proteoglycan associated with mesenchymal tissues. We have described first high expression of syndecan-2 in mesothelioma cells both on messenger RNA and protein levels, shown also by immunocytochemistry. To the best of our knowledge, beside two pilot attempts with semiquantitative assessment in arbitrary units, the quantitative concentrations of syndecan-2 in pleural fluid have not been measured to date.

In the present study we investigated on a large cohort (n=247) the possible additional value of effusion cholesterol and CEA assays to pleural fluid cytology—in particular, in inconclusive cases. Diagnostic utility of syndecan-2 concentrations was also assessed in a subset of these effusions (n=179). The presence of pleural malignancy was established with histology using immunohistochemical panels.

MATERIALS AND METHODS

Patients

In this study we investigated pleural effusions of 247 patients (147 men, aged 22–88 years and 100 women, aged 23–88 years), who were admitted to Korányi National Institute for Pulmonology in Budapest, Hungary. The effusions were submitted for cytological examination. Pleural fluids without additives were prospectively and consecutively collected in different time periods between 2005 and 2019.


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and 2012. Samples were centrifuged for ×600g for 10 min without delay and cell free supernatants were stored at −80°C until assay. Measurements were made regularly when a certain number of samples have accumulated.

According to inclusion criteria, the definitive diagnosis of malignant involvement of the pleura by mesothelioma or metastatic tumours was established on thoracoscopic or ultrasound guided transthoracic lung/pleural biopsies or resection specimens with histopathology, supported by immunohistochemistry with at least four antibodies. The immunohistochemical panel included the following antibodies: TTF1, p40, CD15 or BerEP4, calretinin, podoplanin, WT1, STAT6, CD34, MIB1, napsin A, synaptophysin, CK20, CDX2, and sometimes other tissue-specific markers, as well. p16 FISH analysis was used occasionally in differentiating malignant mesothelioma and benign mesothelial proliferation. Paramalignant effusions caused by cancers without pleural involvement, that is, without presence of clear malignant cells in the pleural fluid were all based on histopathology of the tumour, negative pleural radiology (CT, MRI, positron emission tomography), direct pleural biopsy with negative histology, or negative necropsy within 1 year from sampling. Patients with non-malignant disease caused pleural effusions were followed up for at least a year and excluded if any type of malignancy was diagnosed or they deceased without necropsy, within 1 year from sampling. Tuberculous pleural effusion was diagnosed by identification of acid-fast bacteria (Löwenstein culture, Ziehl-Neelsen staining or PCR in the pleural fluid and/or caseous granulomas in pleural biopsy specimen and/or positive QuantiFERON-TB Gold Plus-test in blood with favourable clinical course after antituberculous therapy. Purulent pleurisy and empyema were verified by radiology, and typical symptoms with cough and fever that toned down with antibiotic treatment.

**Effusion cytology**

Conventional cytological examination of pleural effusions was done on Giemsa stained smears and H&E stained cytospin sediments. Occasional immunocytochemical examination was carried out with antibodies as listed above. The evaluation was accomplished by three experienced cytopathologists (JF, EB, MG) using three diagnostic categories: benign (negative), inconclusive (‘borderline’, ‘suspicous for malignancy’) and malignant (positive). When calculating sensitivity, specificity and predictive values, only ‘malignant’ was considered abnormal, because inconclusive cytology was implicated with the ‘negatives’.

**Tumour markers assay**

Aliquots from supernatant were taken for chemical analysis. Total cholesterol was measured enzymatically, using a colorimetric analysis adapted for the autoanalyser system (Technicon RA-100, Tarrytown, New York, USA). CEA was determined by a two-stage sandwich enzyme linked immunosorbent assay, using mouse and goat monoclonal antibodies (T est RA-100, T arrytown, New York, USA). CEA was determined enzymatically, using a colorimetric analysis adapted for the autoanalyser system (T echnicon RA-100, T arrytown, New York, USA). CEA was determined enzymatically, using a colorimetric analysis adapted for the autoanalyser system (T echnicon RA-100, T arrytown, New York, USA). CEA was determined enzymatically, using a colorimetric analysis adapted for the autoanalyser system (T echnicon RA-100, T arrytown, New York, USA). CEA was determined enzymatically, using a colorimetric analysis adapted for the autoanalyser system (T echnicon RA-100, T arrytown, New York, USA). CEA was determined enzymatically, using a colorimetric analysis adapted for the autoanalyser system (T echnicon RA-100, T arrytown, New York, USA).

**Statistical analysis**

Cut-off limits for chemical analyses were calculated by receiver operating characteristics (ROC), maximised for diagnostic efficacy. The independence of the true diagnosis and the classification by the test used was checked by χ² statistics at a rejection value of p<0.05.

**Table 1 Aetiology of pleural effusions**

<table>
<thead>
<tr>
<th>Pleural effusions</th>
<th>Total</th>
<th>With neoplastic pleural involvement</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total</td>
<td>247</td>
<td>126</td>
</tr>
<tr>
<td>Benign</td>
<td>95</td>
<td>0</td>
</tr>
<tr>
<td>Non-inflammatory</td>
<td>58</td>
<td>0</td>
</tr>
<tr>
<td>Congestive heart failure</td>
<td>42</td>
<td>0</td>
</tr>
<tr>
<td>Liver cirrhosis</td>
<td>6</td>
<td>0</td>
</tr>
<tr>
<td>Irritation, others*</td>
<td>9</td>
<td>0</td>
</tr>
<tr>
<td>Nephrotic syndrome</td>
<td>1</td>
<td>0</td>
</tr>
<tr>
<td>Inflammatory</td>
<td>37</td>
<td>0</td>
</tr>
<tr>
<td>Purulent pleurisy/empyema</td>
<td>21</td>
<td>0</td>
</tr>
<tr>
<td>Tuberculous pleurisy</td>
<td>16</td>
<td>0</td>
</tr>
<tr>
<td>Malignancy-associated</td>
<td>126</td>
<td>26</td>
</tr>
<tr>
<td>Carcinomas</td>
<td>130</td>
<td>24</td>
</tr>
<tr>
<td>Lung</td>
<td>100</td>
<td>19</td>
</tr>
<tr>
<td>Adenocarcinoma</td>
<td>57</td>
<td>9</td>
</tr>
<tr>
<td>Squamous cell</td>
<td>20</td>
<td>6</td>
</tr>
<tr>
<td>Large cell</td>
<td>7</td>
<td>0</td>
</tr>
<tr>
<td>Small cell</td>
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<td>4</td>
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<tr>
<td>Breast</td>
<td>10</td>
<td>0</td>
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<tr>
<td>Colorectal</td>
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<td>1</td>
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<tr>
<td>Ovarian</td>
<td>4</td>
<td>0</td>
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<tr>
<td>Gastric</td>
<td>2</td>
<td>1</td>
</tr>
<tr>
<td>Cervical</td>
<td>2</td>
<td>0</td>
</tr>
<tr>
<td>Pancreatic</td>
<td>1</td>
<td>0</td>
</tr>
<tr>
<td>Renal</td>
<td>1</td>
<td>0</td>
</tr>
<tr>
<td>Prostatic</td>
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<td>0</td>
</tr>
<tr>
<td>Gall bladder</td>
<td>1</td>
<td>0</td>
</tr>
<tr>
<td>Uterine choriocarcinoma</td>
<td>1</td>
<td>0</td>
</tr>
<tr>
<td>Malignant mesothelioma</td>
<td>17</td>
<td>0</td>
</tr>
<tr>
<td>Other malignancies†</td>
<td>5</td>
<td>2</td>
</tr>
</tbody>
</table>

Neoplastic cases are listed also by presence or absence of pleural involvement, and proportion of them producing carcinoembryonic antigen (≥5 ng/mL) is shown in parentheses.

†Pleural thickening: two cases.

‡Lymphoma: three cases, melanoma: two cases.

**RESULTS**

**Patient characteristics**

Aetiology of the effusions is shown in table 1. Of the 247 patients, 95 showed no sign of neoplastic disease, the cause for the benign pleural fluid being partly non-inflammatory consisting of congestive heart failure (42 cases), liver cirrhosis (six cases), pleural irritation comprising also pancreatitis and pleural thickening (nine cases) and one nephrotic syndrome.

Benign inflammatory effusions involved purulent pleurisy and empyema (21 cases), and tuberculous pleurisy (16 cases). The remaining 152 patients had a malignancy. Of 126 patients with malignant involvement of the pleura, 106 had a carcinoma accompanied by pleural carcinomatosis, 17 had mesothelioma and three had other malignancies: two lymphomas and one...
Cytology:
Malignant, $n=88$

Cytology:
Inconclusive, $n=53$

Cytology:
Benign, $n=106$

Neoplastic pleural involvement:

<table>
<thead>
<tr>
<th>No</th>
<th>Yes</th>
</tr>
</thead>
<tbody>
<tr>
<td>121</td>
<td>126</td>
</tr>
</tbody>
</table>

Figure 1  Effusion cytological evaluation ($n=247$) versus neoplastic involvement of the pleura determined by histology ($n=126$).

Of the 106 cases classified by effusion cytology as benign (negative), 104 were correctly identified as non-neoplastic with regard to pleural involvement of a tumour, but two were false negative (figure 1). In 20 of these 106 cases, tumours (18 carcinomas, one melanoma and one lymphoma) were found in the lung and/or lymph nodes, without involvement of the pleura. All 88 cases with malignant effusion cytology had neoplastic pleural involvement caused by carcinoma (carcinomatosis), mesothelioma or other malignancies. In 53 cases (21%), the cytological report was inconclusive. Thirty-six of these proved to have malignant pleural involvement (five mesotheliomas and 31 carcinomas) on later biopsy or necropsy, six had carcinoma without carcinomatosis, whereas no tumour was found in the remaining 11 inconclusive cases. In effusion cytology purulent pleurisy was represented with abundance of neutrophil granulocytes and tuberculous pleurisy by pronounced lymphocytosis.

**CEA concentrations in pleural effusions**

The distribution and covariation of CEA and cholesterol levels is shown in figure 2. All the 81 cases with raised CEA concentrations above the 5 ng/mL cut-off value had a carcinoma; the specificity of CEA for carcinoma was 100% (95% CI 96.9 to 100) whereas the sensitivity was 62.3% (95% CI 53.4 to 70.7), regardless of whether the pleura was involved. Although CEA production is the most common in gastrointestinal cancers, it also occurs in various other types of carcinomas, especially adenocarcinomas and, in 81 cases the secretion of CEA was sufficient to raise the values above the cut-off point (table 1). Of the 100 lung carcinomas 62 had elevated CEA levels in effusion, comprising 43 pulmonary adenocarcinomas, seven squamous cell carcinomas, four large cell carcinomas and eight small cell carcinomas. In 13 carcinoma cases with raised CEA concentration there was no carcinomatosis; however, high CEA values were more common in carcinomas with pleural involvement. All 22 non-epithelial neoplasms, comprising 17 mesotheliomas and five other malignancies, had low CEA levels. Diagnostic performance of CEA assay (figure 3A,B) shows that at optimal cut-off point of 5 ng/mL a moderate (54%) sensitivity and a relatively high (89.3%) specificity is obtained for malignant pleural involvement (table 2).

**Cholesterol concentrations in pleural effusions**

While a raised CEA level is seen not only in pleural carcinomatosis, but rather indicates a CEA producing carcinoma, an increased cholesterol concentration in pleural effusion seems to be a sign of neoplastic pleural involvement (figure 2). Beside nearly all carcinomas accompanied with carcinomatosis (105 cases), all the 17 mesotheliomas and three of other malignancies with pleural involvement (two lymphomas and one melanoma) had elevated cholesterol levels. The optimum diagnostic efficacy is obtained with a threshold of 1.21 mmol/L (figure 3C), and a near-by cut-off point is obtained for the cytologically inconclusive cases (figure 3E). At this cut-off point, malignancies with or without pleural involvement can be well enough separated from each other (figure 2), providing a sensitivity of 99.2% and a specificity of 69.4% for carcinomatosis or neoplastic involvement of the pleura (table 2, figure 3A). Raised cholesterol concentrations, however, are also seen in benign effusions, mostly in inflammatory conditions involving the pleura (37 cases), being either purulent or tuberculous pleurisy. The only benign case with elevated cholesterol level and non-inflammatory cytology was a nephrosis syndrome (figure 2).
Simultaneous measurement of CEA and cholesterol in pleural effusions

Simultaneous rise in cholesterol and CEA concentrations in pleural effusion was specific for carcinomatosis (figure 2), and this combination add diagnostic value (table 2) by increasing the sensitivity of the specific diagnosis of carcinomatosis from 69.8% with cytology alone to 84.1%. In 16 of the 31 carcinomatosis cases with inconclusive effusion cytology, both markers exceeded the respective cut-off values, indicating neoplastic pleural involvement caused by a carcinoma, that is, carcinomatosis (figure 2B). This yields a 51.6% (95% CI 33.1 to 69.9) diagnostic sensitivity among inconclusive cases. Five cases with inconclusive cytology had raised CEA values alone, suggestive of a carcinoma not yet accompanied by carcinomatosis. All five mesotheliomas with borderline effusion cytology and no substantial admixture of inflammatory cells had elevated cholesterol levels, while the two purulent pleurisy cases with inconclusive cytology had both raised cholesterol values and presence of numerous neutrophils in effusion sediment. Among the cases with negative effusion cytology (n=106), the two false negatives, which turned out to have pleural carcinomatosis, have been detected by the simultaneous increase in cholesterol and CEA (figure 1).

Cholesterol concentration combined with assessment of inflammatory cells in effusion sediment

Figure 3A shows ROC curves of cholesterol assay of the total material (n=247) compared with subpopulation without pleural inflammation (n=210). The latter gives at the same threshold magnificent diagnostic performance with high efficacy (99%), separating almost completely malignancies with and without pleural involvement (figures 2 and 3D, table 2). This implies that elevated cholesterol levels, in combination with non-inflammatory effusion cytology, indicate malignant involvement of the pleura, comprising also mesotheliomas and other malignancies, and have recognised all 36 tumour-involved pleura cases with inconclusive cytology.

Syndecan-2 concentrations in pleural effusions

Figure 4 shows distribution of syndecan-2 concentrations in pleural fluids. There were detectable levels exceeding 0.9 ug/mL in all effusions, with no obvious separation between different pathological conditions (see also figure 3A). High levels also occurred in benign non-inflammatory conditions, of which a nephrosis syndrome had the highest value. The highest median value was obtained by the relatively small mesothelioma group with all concentrations exceeding 10 ug/mL. Comparison of syndecan-2 levels in the whole cohort (n=179) with that of the subpopulation with inconclusive effusion cytology cases (n=29, not shown) does not seem to add any distinctive information.

DISCUSSION

Occurrence of tumour cells in effusion sediment can be seen only in neoplastic involvement of the pleura, but the sensitivity of detecting such cells by cytology is restricted, owing to inadequate fluid volume with low cellular yield or bad conservation of cells. Another problem is that benign mesothelial cells may show signs of growth stimulation, that can mimic malignancy, and the induced ‘mesotheliosis’ is sometimes difficult to distinguish from tumour cells by routine cytology alone. These issues
Table 2  Diagnostic values of cytology, cholesterol, carcinoembryonic antigen (CEA), and their combined use in distinguishing between 126 pleural effusions from patients with malignant involvement of the pleura and 121 cases without neoplastic pleural involvement

<table>
<thead>
<tr>
<th>Assays</th>
<th>Definition of abnormality</th>
<th>Sensitivity %, (n/N)</th>
<th>Specificity %, (n/N)</th>
<th>Efficacy %, (n/N)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>(95% CI)</td>
<td>(95% CI)</td>
<td>(95% CI)</td>
</tr>
<tr>
<td>Cytology</td>
<td>Malignant cytology*</td>
<td>69.8 (89/126)</td>
<td>100.0 (121/121)</td>
<td>84.6 (209/247)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>(61.0 to 77.7)</td>
<td>(97.0 to 100.0)</td>
<td>(79.5 to 88.9)</td>
</tr>
<tr>
<td>Cholesterol</td>
<td>Raised cholesterol†</td>
<td>99.2 (125/126)</td>
<td>69.4 (84/121)</td>
<td>84.6 (209/247)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>(95.7 to 100.0)</td>
<td>(60.4 to 77.5)</td>
<td>(79.5 to 88.9)</td>
</tr>
<tr>
<td>CEA</td>
<td>Raised CEA‡</td>
<td>54.0 (68/126)</td>
<td>89.3 (108/121)</td>
<td>71.3 (176/247)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>(44.9 to 62.9)</td>
<td>(82.3 to 94.2)</td>
<td>(65.2 to 76.8)</td>
</tr>
<tr>
<td>Combined cholesterol and CEA</td>
<td>Raised cholesterol and CEA</td>
<td>53.2 (67/126)</td>
<td>100.0 (121/121)</td>
<td>76.1 (188/247)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>(44.1 to 62.1)</td>
<td>(97.0 to 100.0)</td>
<td>(70.3 to 81.3)</td>
</tr>
<tr>
<td>Cytology and/ or combined cholesterol and CEA</td>
<td>Malignant cytology* or raised cholesterol and CEA</td>
<td>84.1 (106/126)</td>
<td>100.0 (121/121)</td>
<td>91.9 (227/247)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>(76.6 to 90.0)</td>
<td>(97.0 to 100.0)</td>
<td>(87.8 to 95.0)</td>
</tr>
<tr>
<td>Cholesterol without inflammation</td>
<td>Raised cholesterol and non-inflammatory cytology§</td>
<td>99.2 (125/126)</td>
<td>98.8 (83/84)</td>
<td>99.0 (208/210)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>(95.7 to 100.0)</td>
<td>(93.5 to 100.0)</td>
<td>(96.6 to 99.9)</td>
</tr>
</tbody>
</table>

* Malignant diagnoses: ‘positives’ only, inconclusive cytology incorporated with ‘negatives’.  †Cut-off point for cholesterol: 1.21 mmol/L.  ‡Cut-off point for CEA: 5 ng/mL. §Non-inflammatory cytology: no substantial presence of neutrophils or lymphocytes in effusion sediment.  CI, confidence interval.

be tracked in blood-serum and effusions.11,12 We have found raised effusional CEA levels beside adenocarcinomas in a few squamous cell, large cell and small cell carcinomas of the lung, which is in line with increased serum levels described in the literature.29,30 Elevated CEA concentrations in effusions have been described in carcinomas with1,2,3,4,13–16 and without carcinoma.16,31–34 The underlying mechanism in the latter case, that is, in paramalignant effusion, is most often due to massive medastinal lymph node metastases. In the present study 13 carcinoma cases without pleural carcinomatosis had increased effusional CEA values (figure 2), and five of them had inconclusive changes on effusion cytology. These morphological changes can be interpreted by non-malignant growth stimulation and proliferation, although case of macroscopically unrevealed metastases35 cannot be ruled out.

Elevated concentration of cholesterol in effusions is more specifically related to exudates, caused by neoplastic or inflammatory involvement of the serous membrane. In neoplastic process cholesterol may originate from cell membranes or cytosol, as a result of disintegration of tumour cells and/or nearby benign serosal cells.12,26 It can also derive from the plasma and interstitial space, as a result of vascular leakage caused by obstructed lymph vessels and/or increased permeability of the affected serous membrane.12,37 One source of cholesterol in inflammatory effusion may be degeneration of leucocytes, that is, the prima facie inflammatory cells, which contain large quantities of this lipid.12,38 Disintegration of serosal cells and/or increased permeability can also occur in massive inflammatory conditions involving the serous membrane,12,37 but these are accompanied with abundant presence of neutrophils or lymphocytes in the effusion sediment, that can easily be detected by cytology. High cholesterol levels in this manner, in combination with non-inflammatory effusion cytology, are therefore most common in association with nonplastic pleural involvement, as represented by high sensitivity and specificity (figures 2 and 3A,D, table 2).

Diagnosis of a malignant condition in the pleural cavity needs maximal specificity to be useful in the clinical praxis. Effusion cytological examination is the first line monitoring that yields a correct diagnosis in most cases with carcinomatosis or neoplastic pleural involvement, however, some samples are reported as inconclusive. Cholesterol value in pleural effusion over 1.21 mmol/L is ‘alone’ a sensitive, though not entirely specific marker for carcinomatosis or neoplastic pleural involvement, while CEA levels exceeding 5 ng/mL can mark a carcinoma. These two chemical assays measure different effects of tumours, such as CEA production versus cell disintegration, and when they were combined, simultaneous increases in both parameters were specific for carcinomatosis, improving the sensitivity of ‘single’ effusion cytology from 69.8% to 84.1% (figure 2, table 2). By this means the correct diagnosis could be established in a number (16/31) of carcinomatosis cases with inconclusive effusion cytology. Measurement of both cholesterol and CEA concentrations in pleural fluid thus adds information to the cytological diagnosis, although elevated cholesterol level solely, with complementary information concerning presence or absence of substantial number of inflammatory cells in effusion sediment overwrites this, providing a superb tool for indication of neoplastic pleural engagement with 99% diagnostic efficacy (figures 2 and 3A and D). Thereby all 36 tumour-involved pleura cases with inconclusive effusion cytology were recognised, comprising also mesotheliomas. The use of cholesterol as a single chemical parameter can primarily be applied or at least interpreted by cytopathologists, because they see both the accidental admixture of abundant number of inflammatory cells in

effusion sediments for excluding these cases, and the occurrence of inconclusive, ‘borderline’, ‘suspicious for malignancy’ cells.

**CONCLUSIONS**

The results of the present study show, that quantitative measurement of syndecan-2 in pleural fluid, although gave detectable concentrations in all effusions with highest median value in mesotheliomas, was non-discriminative between different pathological conditions.

Simultaneous measurement of CEA and cholesterol concentrations in pleural effusion, or at least cholesterol alone, in combination with non-inflammatory fluid cytology, provides additional specific information about neoplastic pleural involvement, and can therefore be used as an adjunct to cytology, above all, in inconclusive cases. This renders an effective assistance primarily for cytopathologists, thereby evading the need for another thoracocentesis.

**Correction notice** This article has been corrected since it was published Online First to resolve formatting errors.

**Handling editor** Tahir S Pillay.

**Acknowledgements** The authors are indebted to Dr Éva Badár for doing cytological diagnostics and her kind cooperation.

**Contributors** All authors (MG, JF, ADK, MM) were involved in the conception and/or design of the study. JF and MG participated in the collection or generation of the study data. MM contributed with laboratory analysis, reagents and tools. ADK was lead statistician of the project, reviewed and provided input for the reporting of the results. MG led the design and initiation of the project and served as a project lead.

**Take home messages**

- Effusional carcinoembryonic antigen (CEA) concentration over 5 ng/mL indicates carcinoma, regardless of pleural involvement.
- Cholesterol level over 1.21 mmol/L marks neoplastic or inflammatory pleural involvement.
- Combined CEA and cholesterol assays increase sensitivity of cytology for carcinoma.
- Elevated cholesterol concentration with non-inflammatory cytology indicates neoplastic pleural involvement.
supervisor throughout. MG, MM, ADK and JF were involved in the analyses and/or interpretation of the data. All authors read, reviewed and approved the present manuscript.

**Funding** This study was supported in part by grants from the Hungarian Kidney Foundation and Research Fund of Uppsala County Council (ALF), Sweden.

**Competing interests** None declared.

**Patient consent for publication** Not required.

**Ethics approval** This manuscript reports a retrospective study of routinely collected samples conducted under approval of Korányi National Institute for Pulmonology (227/2004-IV/1).

**Provenance and peer review** Not commissioned; externally peer reviewed.

**Data sharing statement** Data are available upon reasonable request.

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